

<b>BUSTER reference card</b>	
<b>set up</b>	
Set up for csh or tesh	<code>source /some/where/buster/installed/setup.csh</code>
Set up for sh, bash, ksh, or dash	<code>./some/where/buster/installed/setup.sh</code>
Check all 3rd party tools used by BUSTER work	<code>checkdeps</code>
Download PDB entry (code labc) and convert SF to mtz	<code>fetch_PDB labc</code>
<b>BUSTER refinement most useful options</b>	
Brief help message	<code>refine -h</code>
Default refinement: long 5 big cycles * 100 small cycles	<code>refine -p protein.pdb -m reflects.mtz \ -d res.dir &gt; results.log</code>
No refinement, just maps	<code>refine -p pdb -m mtz -d dir -M MapOnly &gt; log</code>
Quick refine (after coot rebuild)	<code>refine -p pdb -m mtz -d dir -M ShortRunVoid &gt; log</code>
Medium length refinement	<code>refine -p pdb -m mtz -d dir -nbig 2 &gt; log</code>
Use NCS restraints (LSSR)	<code>refine -p pdb -m mtz -d dir -autoncs &gt; log</code>
TLS basic mode	<code>refine -p pdb -m mtz -d dir -M TLSbasic &gt; log</code>
Ligand dictionary (see below)	<code>refine -p pdb -m mtz -d dir -l grade-XXX.cif &gt; log</code>
Use QM method for ligand LIG	<code>refine -p pdb -m mtz -d dir -l grade-LIG.cif -qm LIG \ &gt; log</code>
Rigid body for first big cycle	<code>refine -p pdb -m mtz -d dir -RB &gt; log</code>
Water updating	<code>refine -p pdb -m mtz -d dir -WAT &gt; log</code>
Target restraints (LSSR) to a reference structure	<code>refine -p pdb -m mtz -d dir -target high_res.pdb &gt; log</code>
Occupancy refinement	<code>pdb2occ -p pdb -o pdb-occ.Gelly refine -p pdb -m mtz -d dir -Gelly pdb-occ.Gelly &gt; log</code>
Find ligand binding sites for rhofit	<code>refine -p pdb -m mtz -d dir -L &gt; log</code>
Add hydrogen atoms to protein and its ligand, then refine. Use at better than 2.0Å resolution	<code>hydrogenate -p prot.pdb -l grade-XXX.cif -o protH.pdb refine -p protH.pdb -m mtz -d dir -l grade-XXX.cif &gt; log</code>
An example: initial very long refinement of molecular replacement solution	<code>refine -p MR.pdb -m refl.mtz -d res.dir \ -RB -target MR.pdb -nbig 10 &gt; results.log</code>
<b>Looking at BUSTER refinement results</b>	
Produce report on <code>refine</code> run	<code>buster-report -d dir</code>
view buster-report output	<code>firefox dir-report/index.html</code>
Start <code>coot</code> to see structure with maps plus “unhappy atoms” list	<code>visualise-geometry-coot res.dir</code>

For latest version of BUSTER reference card see file `$BDG_home/docs/buster_reference_card.pdf`

<b>BUSTER reference card</b>	
<b>cif restraint dictionary preparation</b>	
Brief help message	<code>grade -h</code> and <code>grade_PDB_ligand -h</code>
SMILES residue name LIG note: use of ' '	<code>grade -resname LIG 'C1CN(CCN1CCO)CCS(=O)(=O)O'</code>
From mol2 file (with hydrogen atoms)	<code>grade -in ligand.mol2 -resname LIG</code>
Charged ligand	<code>grade -in acid.mol2 -resname LIG -charge -1</code>
Ligand exists in PDB eg 3AS	<code>grade_PDB_ligand 3AS</code>
Examine (and edit) cif dictionary	<code>EditREFMAC grade-LIG.cif grade-CUF.pdb LIG</code>
<b>Ligand fitting</b>	
Brief help message	<code>rhofit -h</code>
Fit ligand XXX	<code>rhofit -p protein.pdb -m refine.mtz -l grade-XXX.cif -d rhofit.dir</code>
Fit ligand in 2 sites	<code>rhofit -p prot.pdb -m mtz -l cif -d dir -xclusters 2</code>
Allow chiral centres to invert during fit	<code>rhofit -p prot.pdb -m mtz -l cif -d dir -nochirals</code>
Look at <b>rhofit</b> results using <b>coot</b>	<code>cd rhofit.dir</code> <code>visualise-rhofit-coot</code>
<b>refine</b> from rhofit best fits in each site (use with caution!)	<code>refine -p rhofit.dir/merged.pdb \</code> <code>-l rhofit.dir/best.cif -m mtz -autoncs &gt; log</code>

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